



Precision Medicine and Mysteries in Clinical Microbiology: Rationalizing Epidemiology, Genotype, and Phenotype To Guide Therapeutics

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ABSTRACT Whole-genome sequencing (WGS) using MinION was used to characterize high-risk clones of *Escherichia coli* and *Klebsiella pneumoniae* harboring *bla*_{NDM-5}, *bla*_{OXA-181}, and *bla*_{CTX-M-15}, as well as *Pseudomonas aeruginosa* harboring *bla*_{NDM-1} in a patient who received health care in India. Synergy testing demonstrated the activity of aztreonam and ceftazime-avibactam in combination. This case illustrates a “precision medicine” approach where deeper understanding of the genotype through WGS and of the phenotype through synergy testing formed the basis for rational combination therapy.

Khan et al. describe the fascinating case of a woman who, after receiving medical care in India, developed infection with Gram-negative pathogens resistant to carbapenems (1). Additionally, she was colonized with *Candida auris*, an emerging candidal species characterized by resistance to azoles. Cultures of the urine grew *Escherichia coli*, demonstrated by PCR to harbor *bla*_{NDM-5}, *bla*_{OXA-181}, and *bla*_{CTX-M-15}, as well as *Klebsiella pneumoniae* containing *bla*_{NDM-5} and *bla*_{CTX-M-15}; culture of a sputum sample grew *Pseudomonas aeruginosa* harboring *bla*_{NDM-1}. Guided by this knowledge, clinicians treated her with aztreonam, which is stable in the presence of the metallo-β-lactamase NDM, in conjunction with avibactam (administered as ceftazidime-avibactam), which protects aztreonam from inactivation by CTX-M-15 and OXA-181 β-lactamases. Demonstration of *in vitro* synergy of this combination of antibiotics affirmed their choice of therapy. The patient, whose infection did not respond to colistin despite the *in vitro* activity of this agent, appeared to have a favorable clinical response to aztreonam and ceftazidime-avibactam, although she eventually died after a stroke.

This case features the use of whole-genome sequencing (WGS) to determine bacterial and plasmid sequence types, as well as the resistome of *E. coli*, *K. pneumoniae*, and *P. aeruginosa*. Specifically, long-read genomic data were procured with the MinION nanopore sequencer, with additional complementary short-read sequences obtained using Illumina. MinION is a portable and affordable device that permits, for instance, real-time genomic analysis for the rapid diagnosis, detection of antibiotic resistance, and surveillance of *Mycobacterium tuberculosis* (2). The use of MinION provided insights that, in conjunction with other microbiological and pharmacological information, were

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clinically useful and herald a “precision medicine” approach for the treatment of Gram-negative bacterial infections (3).

The strain of *E. coli* identified in this case, sequence type 167 (ST167), usually harbors *bla*_{NDM-5} or *bla*_{OXA-181} within a plasmid of the IncX3 type, which can also be hosted by other sequence types of *E. coli* and other bacterial species (4). The IncX3 plasmid found in this isolate did not contain carbapenemase genes but contained the cephalosporinase *bla*_{CTX-M-15}. Rather, *bla*_{NDM} was in an IncF plasmid surrounded by a genetic environment like that of other *bla*_{NDM} plasmids in *E. coli* from Asia and Europe (5). Similarly, *K. pneumoniae* ST16, also isolated in this case, has been reported harboring *bla*_{NDM-5} in Asia and Europe. Variability in the plasmid and resistance content of *K. pneumoniae* ST16 suggests that different variants of that clone circulate worldwide (6). *P. aeruginosa* harboring *bla*_{NDM1} found in this case, is not uncommon and was reported recently in association with ST773 (7).

All these sequence types are deemed “international high-risk clones” and appear to be stable platforms for the maintenance of carbapenemase genes. High-risk clones seem to have a propensity to acquire plasmids and undergo genetic rearrangements that facilitate horizontal transmission of antibiotic resistance genes and, as in this case, disseminate across continents (8). Although the exact interplay between high-risk clones, plasmids, and other mobile genetic elements is not clear, it must serve the process of evolution and permit bacterial adaptation to conditions in the environment, including antibiotic selective pressure.

This case is extraordinary because of the simultaneous infection with *E. coli*, *K. pneumoniae*, and *P. aeruginosa* harboring multiple carbapenemases, as well as colonization with *C. auris*. Despite genetic details obtained through WGS, it remains impossible to reconstruct how this occurred. Whether she became colonized inside or outside the hospital in India where she was initially treated, or even plausibly in the rehabilitation facility in the United States where she was recovering, remains unknown. Also, recapitulating the type, duration, and sequence of antibiotics she received is not feasible, and therefore the evolutionary path by which resistance emerged cannot be retraced (9). Clarifying those details could recast this patient’s unlikely misfortune as the logical culmination of acquisition, rearrangement, and dissemination of mobile genetic elements and high-risk clones in the context of environmental exposure, antibiotic selective pressure, and international travel. Furthermore, it would point toward strategies for the prevention and control of such events.

The promise of precision medicine is that a deeper understanding of the mechanisms of disease can be a compass pointing toward better therapies. The rationale for the use of aztreonam and ceftazidime-avibactam was grounded on the hydrolytic profiles of each of the enzymes against aztreonam and the ability of avibactam to inhibit key β -lactamases (10). This knowledge supported the use of a combination containing aztreonam and avibactam but was insufficient in the absence of information about the bacterial phenotype garnered through more conventional microbiologic methods. Therefore, demonstration through antimicrobial susceptibility testing of *in vitro* activity of aztreonam against *P. aeruginosa* and synergy between ceftazidime-avibactam and aztreonam against *E. coli* and *K. pneumoniae* was essential.

Similarly, colistin appeared active *in vitro* against all the pathogens, yet this polymyxin failed in achieving a clinical response. This underscores the notion that colistin has serious shortcomings for the treatment of infections caused by Gram-negative bacteria. The evidence documenting how and why colistin fails is robust: antibiotic concentrations are insufficient at the site of infection, the therapeutic window is narrow with frequent nephrotoxicity, and resistance can emerge during treatment (11). Conversely, there are unexplored aspects of how and why the combination of aztreonam and ceftazidime-avibactam is effective against metallo- β -lactamase-harboring bacteria, such as optimal timing and dosing, antibiotic exposures in different compartments, and the adjuvant role of ceftazidime targeting penicillin-binding proteins. Answering these and other questions, and proving the efficacy and safety of this combination in well-designed clinical trials, should be the aim of a conscientious research program.

Ultimately, the use of genotypic and phenotypic data in this case mattered because it led to the selection of an effective antibiotic combination against carbapenemase-producing bacteria where the usual therapy failed. This was possible because of progress in the basic science of resistance mechanisms and investments in the development of new antibiotics. Even though this case is of interest to the community of scientists and clinicians who have access to these tools, it remains to be seen whether precision medicine as illustrated here increases quality, safety, and value and if it is affordable and universally applicable. The latter aspect is crucial, considering that the burden of carbapenemase-mediated resistance is highest in regions of the world with low incomes. By improving access to WGS, platforms such as the MinION sequencer may help upend diagnostic and treatment paradigms, which could be transformative in the global fight against antibiotic resistance.

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REFERENCES

1. Khan A, Shropshire WC, Hanson B, Dinh AQ, Wanger A, Ostrosky-Zeichner L, Arias CA, Miller WR. 2020. Simultaneous infection with *Enterobacteriaceae* and *Pseudomonas aeruginosa* harboring multiple carbapenemases in a returning traveler colonized with *Candida auris*. *Antimicrob Agents Chemother* 64:e01466–19. <https://doi.org/10.1128/AAC.01466-19>.
2. Votintseva AA, Bradley P, Pankhurst L, Del Ojo Elias C, Loose M, Nilgiriwala K, Chatterjee A, Smith EG, Sanderson N, Walker TM, Morgan MR, Wyllie DH, Walker AS, Peto TEA, Crook DW, Iqbal Z. 2017. Same-day diagnostic and surveillance data for tuberculosis via whole-genome sequencing of direct respiratory samples. *J Clin Microbiol* 55:1285–1298. <https://doi.org/10.1128/JCM.02483-16>.
3. Perez F, El Chakhtoura NG, Papp-Wallace KM, Wilson BM, Bonomo RA. 2016. Treatment options for infections caused by carbapenem-resistant *Enterobacteriaceae*: can we apply “precision medicine” to antimicrobial chemotherapy? *Expert Opin Pharmacother* 17:761–781. <https://doi.org/10.1517/14656566.2016.1145658>.
4. Mouftah SF, Pál T, Darwish D, Ghazawi A, Villa L, Carattoli A, Sonnevend Á. 2019. Epidemic IncX3 plasmids spreading carbapenemase genes in the United Arab Emirates and worldwide. *Infect Drug Resist* 12:1729–1742. <https://doi.org/10.2147/IDR.S210554>.
5. Giufrè M, Errico G, Accogli M, Monaco M, Villa L, Distasi MA, Del Gaudio T, Pantosti A, Carattoli A, Cerquetti M. 2018. Emergence of NDM-5-producing *Escherichia coli* sequence type 167 clone in Italy. *Int J Antimicrob Agents* 52:76–81. <https://doi.org/10.1016/j.ijantimicag.2018.02.020>.
6. Espinal P, Nucleo E, Caltagirone M, Mattioni Marchetti V, Fernandes MR, Biscaro V, Rigoli R, Carattoli A, Migliavacca R, Villa L. 2019. Genomics of *Klebsiella pneumoniae* ST16 producing NDM-1, CTX-M-15, and OXA-232. *Clin Microbiol Infect* 25:385.e1–385.e5. <https://doi.org/10.1016/j.cmi.2018.11.004>.
7. Kocsis B, Toth A, Gulyas D, Ligeti B, Katona K, Rokusz L, Szabo D. 2019. Acquired qnrVC1 and blaNDM-1 resistance markers in an international high-risk *Pseudomonas aeruginosa* ST773 clone. *J Med Microbiol* 68:336–338. <https://doi.org/10.1099/jmm.0.000927>.
8. Mathers AJ, Peirano G, Pitout J. 2015. The role of epidemic resistance plasmids and international high-risk clones in the spread of multidrug-resistant *Enterobacteriaceae*. *Clin Microbiol Rev* 28:565–591. <https://doi.org/10.1128/CMR.00116-14>.
9. Nichol D, Jeavons P, Fletcher AG, Bonomo RA, Maini PK, Paul JL, Gatenby RA, Anderson ARA, Scott JG. 2015. Steering evolution with sequential therapy to prevent the emergence of bacterial antibiotic resistance. *PLoS Comput Biol* 11:e1004493. <https://doi.org/10.1371/journal.pcbi.1004493>.
10. Marshall S, Hujer AM, Rojas LJ, Papp-Wallace KM, Humphries RM, Spellberg B, Hujer KM, Marshall EK, Rudin SD, Perez F, Wilson BM, Wasserman RB, Chikowski L, Paterson DL, Vila AJ, van Duin D, Kreiswirth BN, Chambers HF, Fowler VG, Jacobs MR, Pulse ME, Weiss WJ, Bonomo RA. 2017. Can ceftazidime-avibactam and aztreonam overcome β -lactam resistance conferred by metallo- β -lactamases in *Enterobacteriaceae*? *Antimicrob Agents Chemother* 61:e02243-16. <https://doi.org/10.1128/AAC.02243-16>.
11. Tsuji BT, Pogue JM, Zavascki AP, Paul M, Daikos GL, Forrest A, Giacobbe DR, Viscoli C, Giamarellou H, Karaiskos I, Kaye D, Mouton JW, Tam VH, Thamlikitkul V, Wunderink RG, Li J, Nation RL, Kaye KS. 2019. International consensus guidelines for the optimal use of the polymyxins: endorsed by the American College of Clinical Pharmacy (ACCP), European Society of Clinical Microbiology and Infectious Diseases (ESCMID), Infectious Diseases Society of America (IDSA), International Society for Anti-infective Pharmacology (ISAP), Society of Critical Care Medicine (SCCM), and Society of Infectious Diseases Pharmacists (SIDP). *Pharmacotherapy* 39:10–39. <https://doi.org/10.1002/phar.2209>.